

Customized CNN Model for Multiple Illness Identification in Rice and Maize

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Abstract: Crop diseases imperil global food security and economies, demanding early detection and effective management. Convolutional Neural Networks (CNNs), particularly in rice and maize leaf disease classification, have gained traction due to their automatic feature extraction capabilities. CNN models eliminate manual feature extraction, enabling precise disease diagnosis based on learned features. Researchers have rapidly advanced these models, achieving promising results. Leaf disease characteristics like color changes, texture variations, and lesion appearance have been identified as useful for automated diagnosis using machine learning. Developing CNN models involves crucial stages: dataset preparation, architecture selection, hyperparameter tuning, and model training and evaluation. Diverse and accurately annotated datasets are pivotal, and appropriate CNN architecture selection, such as ResNet101 and XceptionNet, ensures optimal performance. These architectures' pre-training on vast image datasets enhances feature extraction. Hyperparameter tuning fine-tunes the model, and training and evaluation gauge its precision. CNN models hold potential to enhance rice and maize productivity and global food security by effectively detecting and managing diseases.

Keywords: Maize Leaf Disease, Rice Leaf Disease, Nutrient Deficiency, CNN, Fusion.

I. Introduction:

Crop diseases pose a major threat to the global food supply chain, resulting in significant economic losses and jeopardizing food security. Early detection and timely management of crop diseases are crucial to prevent their spread and minimize the damage caused by them. Image-based methods have emerged as a promising tool for detecting and classifying crop diseases accurately and efficiently. Deep learning-based techniques, specifically Convolutional Neural Networks (CNNs), have shown remarkable performance in image classification tasks [1].

In recent years, CNNs have been extensively used for leaf disease classification in various crops, including rice and maize. Leaf diseases in rice and maize are caused by various fungal, bacterial, and viral pathogens, leading to significant crop losses worldwide. Early and accurate detection of leaf diseases in these crops is essential for timely and effective management of these diseases [2].

CNN-based models have demonstrated excellent performance in leaf disease classification tasks in various crops. These models learn discriminative features automatically from the images, eliminating the need for manual feature extraction, which was a significant challenge in traditional image classification methods. The features learned by the CNN models are highly representative of the underlying patterns and

characteristics of the disease, enabling accurate classification and diagnosis [3].

The development of CNN models for rice and maize leaf disease classification has gained significant momentum in recent years [4]. Researchers have proposed various CNN architectures and training strategies to improve the accuracy and efficiency of these models. Several studies have reported promising results in rice and maize leaf disease classification using CNN models, indicating the potential of these models for practical applications.

The changes in leaf features that are typically observed in the presence of leaf diseases such as Gray Spot, Blight, and Measles may vary depending on the specific disease and the plant species being affected. However, here are some common changes that may be observed in these diseases compared to a healthy condition:

1. **Gray Spot:** Gray Spot is a fungal disease that affects many different plant species. In infected leaves, gray or brown spots with yellow halos may appear. The affected areas may also have a fuzzy or powdery appearance due to the growth of fungal spores. Some common changes in leaf features that may be observed in Gray Spot include changes in leaf color and texture, as well as the presence of lesions or spots.

2. **Blight:** Blight is a broad term that refers to any disease that causes rapid and widespread death of plant tissue. Symptoms of blight can include wilting, yellowing, and browning of leaves, as well as the presence of lesions or spots. Some common changes in leaf features that may be observed in blight include changes in leaf color, texture, and shape, as well as the presence of lesions or spots.
3. **Measles:** Measles is a viral disease that affects many different plant species. In infected leaves, small, circular or irregularly shaped spots may appear, which can be either light or dark in color. Some common changes in leaf features that may be observed in Measles include changes in leaf color and texture, as well as the presence of lesions or spots.

Overall, changes in leaf color, texture, and the presence of lesions or spots are common features that may be observed in the presence of leaf diseases such as Gray Spot, Blight, and Measles [5]. These changes can be used to develop machine learning models for automated disease diagnosis and classification. Figure 1 shows the respective diseases impacts on the leaves of selected crops.

The development of CNN models for rice and maize leaf disease classification involves several critical steps, including dataset preparation, model architecture selection, hyperparameter tuning, and model training and evaluation. The choice of dataset and its quality are crucial for the performance of the CNN models. The dataset should be diverse, representative, and annotated accurately to ensure reliable training and evaluation of the models.

The selection of the appropriate CNN architecture is also essential for achieving optimal performance in leaf disease classification tasks. Various CNN architectures, such as VGG, ResNet, Inception, and DenseNet, have been used for rice and maize leaf disease classification, each with its unique advantages and limitations. The architecture should be selected based on the specific requirements of the task and the dataset characteristics.

Hyperparameter tuning is another critical step in CNN model development, which involves optimizing the model's learning rate, batch size, regularization, and other parameters to achieve the best possible performance [6]. Several techniques, such as grid search, random search, and Bayesian optimization, have been used for hyperparameter tuning in CNN model development.

Finally, the model training and evaluation stage involves training the model on the dataset and evaluating its performance on the test set. Various performance metrics, such as accuracy,

precision, recall, and F1 score, are used for evaluating the model's performance. Cross-validation and data augmentation techniques are often used to improve the model's generalization ability and reduce overfitting. The development of CNN models for rice and maize leaf disease classification has shown great promise for accurate and efficient disease detection and management. The advancement of these models is expected to play a vital role in enhancing crop productivity and ensuring global food security.

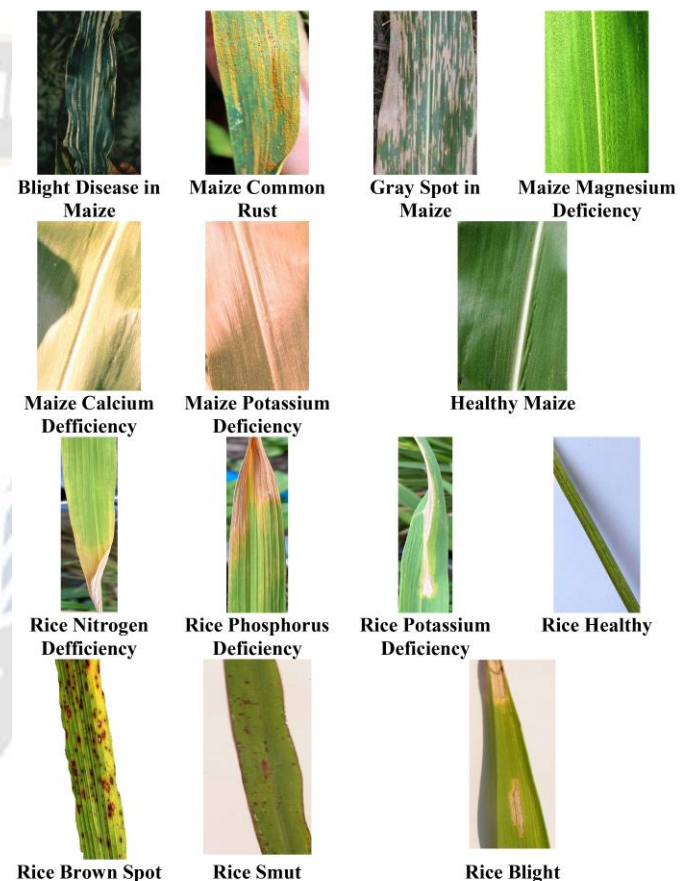


Figure 1: Different Diseases and Healthy Condition Images of Rice and Maize

ResNet101 [7] and XceptionNet [8] are both deep neural network architectures that have been pre-trained on large image datasets, such as ImageNet. These networks have been shown to be highly effective at feature extraction, which makes them well-suited for tasks such as leaf disease classification.

ResNet101 is a deep residual network that is designed to address the problem of vanishing gradients in deep neural networks. It achieves this by using skip connections that allow information to bypass multiple layers and be directly fed into deeper layers. This architecture enables ResNet101 to learn highly discriminative features that are useful for tasks such as leaf disease classification. XceptionNet, on the other hand, is a network architecture that uses a combination of convolutional

layers with different filter sizes and pooling operations to extract features at multiple scales. This allows XceptionNet to capture both fine-grained and coarse-grained features, which can be useful for tasks such as leaf disease classification.

Both ResNet101 and XceptionNet have been shown to be highly effective at feature extraction for a wide range of image classification tasks, including leaf disease classification. This is due to their ability to learn highly discriminative features that are representative of the underlying patterns and structures in the images. Additionally, because they have been pre-trained on large image datasets, they are able to generalize well to new images, which makes them useful for tasks such as transfer learning.

II. Related Work

The recent surge in deep learning, especially Convolutional Neural Networks (CNNs), for plant disease detection has sparked notable research. Abade et al. [9] investigated CNN algorithms, emphasizing trends like PlantVillage and TensorFlow. Dhaka et al. [10] delved into CNN-based techniques, highlighting framework choice, model architecture, and pre-processing's role. Their study underscored dataset quality's impact. Nagaraju et al. [11] reviewed 80+ Deep Learning (DL) disease studies, emphasizing pre-processing's significance. Kamilaris et al. [12] showcased DL's superiority in addressing agricultural challenges, surpassing traditional methods. Fernandez-Quintanilla et al. [13] explored crop weed monitoring, stressing weed control and data collection through diverse monitoring methods, cloud platforms, and accessible information. These studies collectively reveal the transformative potential of deep learning in agriculture. Setiawan et al. [14] provide comparison of machine learning and deep learning for maize leaf disease classification.

Lu et al. [15] evaluated CNN performance in plant disease classification, exploring architectures, strengths, and improvements. Golhani et al. [16] highlighted hyperspectral data's potential for plant disease diagnosis, emphasizing comprehensive data collection. Mosleh et al. [17] developed an effective CNN model for potato disease detection, showcasing high accuracy. Huang et al. [18] introduced DenseNet, enhancing CNN architectures for information flow. Li et al. [19] extended this with fire-FRD-CNN and mobile-FRD-CNN, optimizing feature map generation. Lee et al. [20] simplified disease classification with GoogleNet-BN for Plant Village dataset. Mao et al. [21] optimized models with depth-wise separable convolution and filter pruning. Singh et al. [22] proposed joint pruning and fine-tuning for model efficiency. Li et al. [23] emphasized compact, accurate models through CNNPruner, aligning with efficiency demands. Qian et al. used self-attention mechanism in CNN model for Maize leaf disease

identification. Ma et al. [24] provided transfer learning approach for maize leaf disease detection work.

Rawat et al. [25] used ResNet50 for detection rice leaf disease detection along with 4000 images sample size. Yang et al. [26] used stacking approach in CNN based rice leaf disease detection method. Improvement in AlexNet, ResNet50 and MobileNet-V3 are observed with stacking method. Simhadri et al. [27] provided analysis of 15 different CNN models in transfer learning approach for rice leaf disease detection. Inception-V3 was found effective among all the models with better accuracy of rice leaf disease detection.

Collectively, these studies underscore the dynamic landscape of plant disease detection, driven by the evolution of deep learning models and their application-specific adaptations. The pursuit of accurate, efficient, and scalable disease detection systems remains a driving force in the realm of agricultural technology, with implications for precision agriculture, crop management, and sustainable food production.

This study's limitations encompass its focus on specific neural network architectures and datasets, potentially hindering generalization. The proposed techniques, although effective, may require further optimization. The scope extends to enhancing model efficiency, but does not extensively explore alternative approaches or real-world deployment challenges.

III. Methodology

The important features for leaf disease classification may vary depending on the type of disease and the specific dataset being used. However, some common features that have been found to be useful in leaf disease classification include:

1. **Color features:** Leaf diseases can often cause changes in leaf color, such as yellowing, browning, or spotting. Therefore, color features such as mean color intensity, color histograms, and color channel statistics can be informative for disease classification [28].
2. **Texture features:** Leaf diseases can also cause changes in leaf texture, such as roughness or deformation. Texture features such as local binary patterns, gray level co-occurrence matrices, and wavelet transforms can capture these changes and aid in disease classification.
3. **Shape features:** Leaf diseases can also affect the overall shape and size of the leaf, as well as the shape and size of individual lesions or spots. Shape features such as area, perimeter, circularity, and eccentricity can capture these changes and aid in disease classification.
4. **Symmetry features:** Some leaf diseases can cause asymmetric damage to the leaf, which can be captured by symmetry features such as the difference in shape or texture between the left and right halves of the leaf.

5. Structural features: The underlying structure of the leaf, such as the arrangement of veins or the presence of stomata, can also be informative for disease classification.
6. Contextual features: The context in which the leaf image was taken can also be useful for disease classification. For example, the type of plant, the location of the plant, and the time of year can all affect the likelihood of certain diseases occurring.

These features can be extracted from leaf images using various techniques such as image processing and computer vision algorithms, and can be used to train machine learning models for leaf disease classification.

The leaf disease and nutrient Deficiency identification work is carried out as the block diagram shown in figure 1. The dataset preparation, feature extraction and model training, testing the model is detailed in further sections.

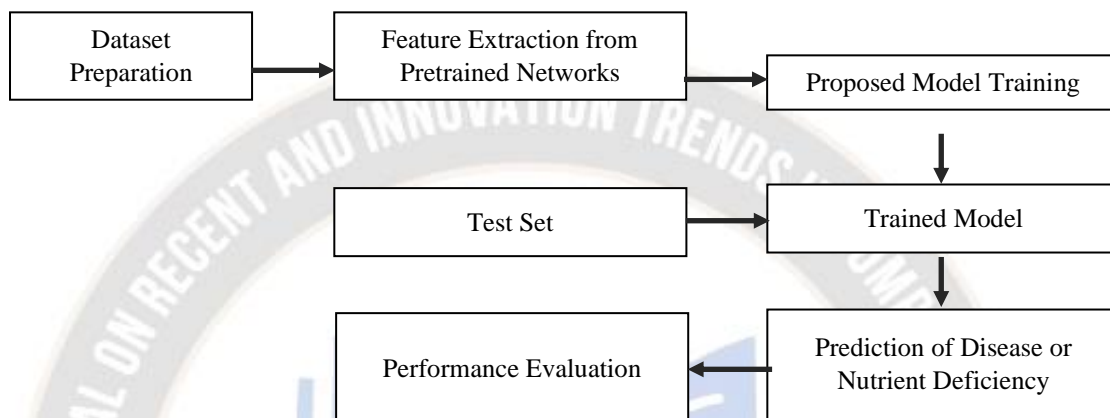


Figure 1: Proposed System Framework

3.1 Dataset Preparation

The dataset of nutrient deficiency in Maize [29] and Rice [30] are collected from standard public repositories as shown in table 1. The images obtained are then passed through augmentation process to get maximum combinations for better training of the models. The respective details are shown in table 1.

Table 1: Dataset Details

Dataset Name	Number of Images (Healthy, Ca, K, Mg)	Images After Augmentation
IPNI	680 (176, 158, 171, 175)	2720
Kaggle Maize Nutrient Dataset	800 (163, 186, 237, 214)	3200
Total Number of Images	1480	5920

Table 2: Maize Leaf Disease Dataset Details

Type of Disease	Images	After Augmentation
Blight	1146	4584
Common Rust	1306	5224
Gray Leaf Spot	574	2296
Healthy	1162	4648
Total Images	4188	16752

Table 3: Maize Nutrient Deficiency Leaf Dataset:

Deficiency	Images	After Augmentation
Calcium	339	1356
Potassium	344	1376
Nitrogen	408	1632
Healthy	389	1556
Total Images	1480	5920

Table 4: Rice Leaf Disease Dataset

Type of Disease	Images	After Augmentation
Brown Spot	523	2092
Hispa	565	2260
Leaf Blast	779	3116
Healthy	1488	5952
Total Images	3355	13420

Table 5: Rice Leaf Nutrient Deficiency Dataset

Deficiency	Images	After Augmentation
Phosphorus	333	1332
Potassium	383	1532
Nitrogen	440	1760
Healthy	389	1556
Total Images	1545	6180

3.2 Feature extraction

3.2.1 Features from Pretrained Network

When using ResNet101 and Xception in parallel for feature extraction, we need to determine the optimal layer from each

network to extract complementary and discriminative features. The choice of layers depends on the specific task and dataset. For leaf disease classification, a good starting point for ResNet101 may be to extract features from the output of the last layer in the last residual block, just before the global average pooling layer. In ResNet101, with consideration of all the sublayers within the Residual Block, the total number of layers is 347. For Xception, a good starting point may be to extract features from a specific layer. To select an appropriate layer in Xception for feature extraction, we can consider the deeper layers closer to the end of the network, as they tend to capture more abstract and high-level features. The specific layer number in Xception will depend on the implementation and

how you define a "layer". Once features have been extracted from ResNet101 and Xception, they can be concatenated and passed through an attention layer. The attention layer can selectively weight the importance of different features based on their relevance to the classification task. Various attention mechanisms, such as channel-wise or spatial attention, can be employed. Finally, the features from the attention layer can be fed into one or more dense layers for the final classification task. The architecture of the dense layers will depend on the specific requirements of the task and dataset, and can be designed accordingly.

3.2.2 Residual Block Based Network

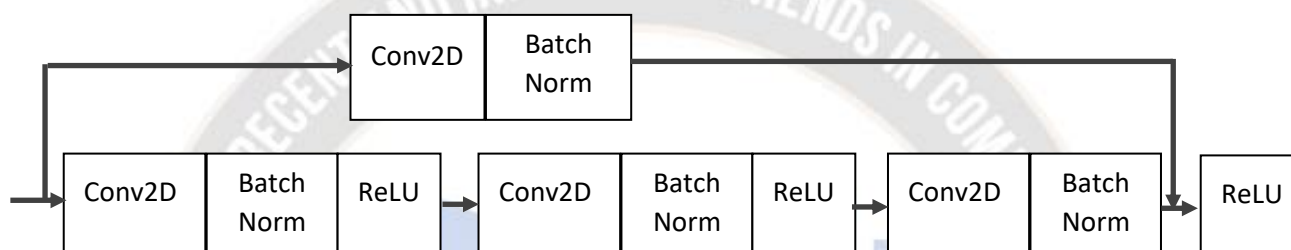


Figure 2: Residual Block

Residual blocks play a crucial role in deep learning models, especially in convolutional neural networks (CNNs) [11]. They were introduced in the paper "Deep Residual Learning for Image Recognition" by He et al. in 2016 and have since become an integral component in numerous cutting-edge models.

At a high level, a residual block enables a neural network to learn residual functions, which capture the difference between the input and output of a layer. This is achieved by incorporating a shortcut connection that allows the network to bypass one or more layers and directly propagate information between layers. The typical architecture of a residual block is depicted in Figure 3.

The process begins with the input passing through a convolutional layer, followed by batch normalization and a Rectified Linear Unit (ReLU) activation. Subsequently, another

convolutional layer and batch normalization are applied, followed by the introduction of the shortcut connection. The shortcut connection combines the input with the output of the second convolutional layer using element-wise addition. The resulting sum is then subjected to another ReLU activation before being returned as the output of the residual block.

The effectiveness of residual blocks can be attributed to the shortcut connection, which allows the network to optimize the residual function instead of solely focusing on mapping the input to the output. This property is particularly advantageous when training deep networks, as it helps address the issue of vanishing gradients that can occur during backpropagation through multiple layers.

In mathematical terms, the representation of a residual block can be written as:

$$Output = activation \left(Convolution \left(normalization \left(activation (Convolution (input) \right) \right) \right) + input \right) \dots(1)$$

Here, input refers to the input of the block, convolution denotes the convolutional operation, normalization represents the batch normalization operation, activation signifies the ReLU activation function, and output indicates the output of the block. The addition operation is performed element-wise, and the resulting output is returned after passing through an activation function.

In summary, the shortcut connection in residual blocks establishes a direct link between the input and output of a layer, facilitating the bypassing of one or more layers. This mechanism enables efficient information transmission within the network and contributes to the success of residual blocks in deep learning models.

3.2.3 Proposed Architecture

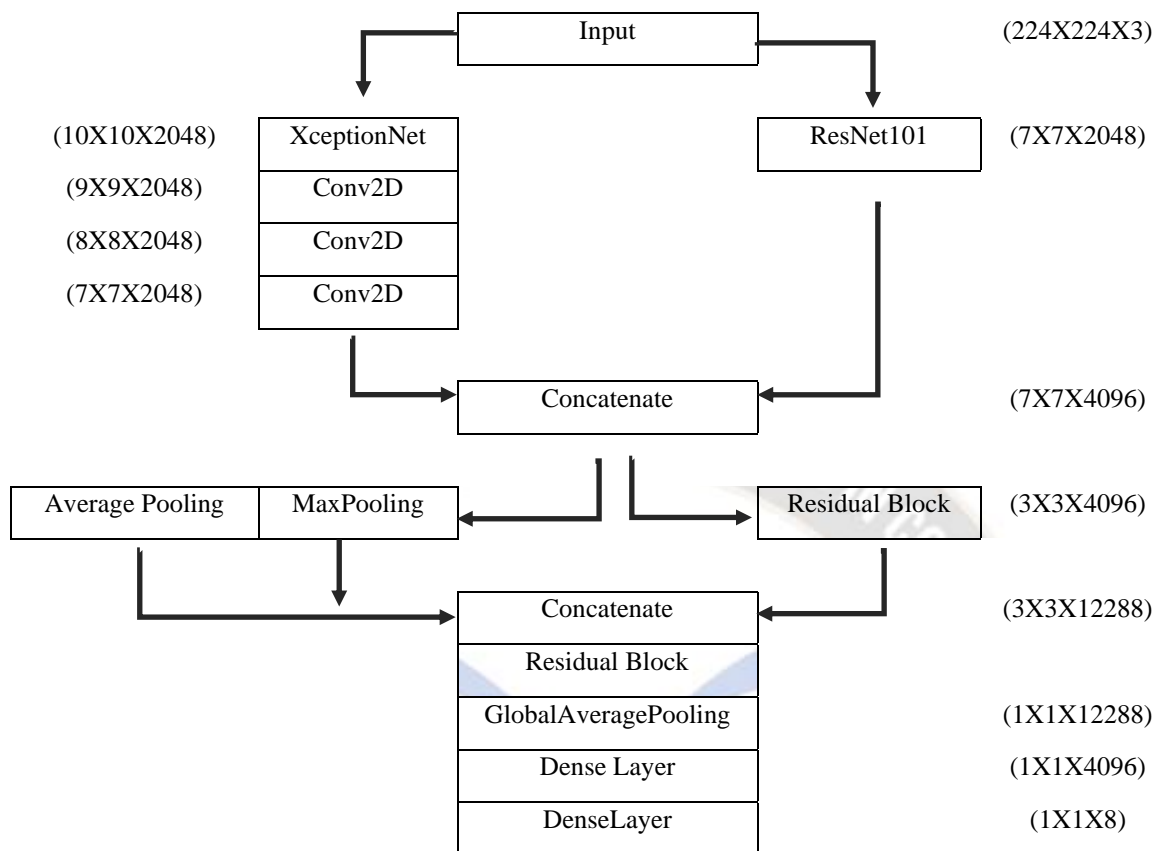


Figure 3: Architecture Proposed Model

The proposed model consists of an input layer that receives an input image with dimensions (224x224x3). The ResNet101 and Xception models are employed to extract features from this image, yielding feature maps with dimensions of (7x7x2048) and (10x10x2048), respectively. These feature maps are obtained from the last selected layer before the Global Average Pooling layer in both models. To match the dimensions of the features without losing valuable information, the output feature map from Xception undergoes a sequence of three convolutional layers, each reducing the dimensions. This ensures compatibility with the dimensions of the feature map from ResNet101. The two sets of feature maps are then concatenated. An attention effect is achieved through the parallel combination of Average Pooling and Max Pooling. The output from the attention layer, along with the parallel output from the Residual Block, is concatenated. The Residual Block employed here is the Residual Pooling block, which reduces the dimensions to 3x3x4096 to match those of the Attention Layer. The output from both sides is concatenated. The features are then passed through another residual block before reaching the Global Average Pooling layer. The output of the Global Average Pooling layer is then fed into a Dense layer with 256 hidden neurons. This Dense layer utilizes the ReLU activation function. Finally, the last Dense layer performs the

classification task using the Softmax activation function, assigning the features to one of the eight classes. The proposed model combines features from ResNet101 and Xception through dimension matching and concatenation. It incorporates an attention layer and Residual Blocks to enhance feature representation. The classification is performed using Dense layers with ReLU activation and concludes with the final Softmax layer for class assignment.

$$ResNet101_{output} = ResNet101(InputImage) \quad \dots(2)$$

where Input_Image is the input image with dimensions (224x224x3).

$$Xception_{output} = Xception(InputImage) \quad \dots(3)$$

where Input_Image is the input image with dimensions (224x224x3).

$$Xception_{output_Conv} = Conv2D(Xception_{output}) \quad \dots(4)$$

where Xception_Output has dimensions (10x10x2048) and Xception_Output_Conv has reduced dimensions.

$$\begin{aligned} & \text{Concatenated}_{Features} \\ & = \text{Concatenate}(\text{ResNet101}_{output}, \text{Xception}_{output}) \end{aligned} \quad \dots(5)$$

where Concatenated_Features combines the features from ResNet101 and Xception.

3.2.4 Attention Layer:

Average Pooling: The Average Pooling operation calculates the average value of each local region in the input. It computes the average of the values within a window or kernel, sliding it across the input feature map. Average Pooling helps capture the overall distribution and general patterns in the data.

Max Pooling: The Max Pooling operation, on the other hand, selects the maximum value from each local region in the input. It identifies the most prominent or salient features within each kernel window. Max Pooling is effective in capturing the most distinctive and important features. By combining Average Pooling and Max Pooling in parallel, the attention layer can gather information from both average-level and maximum-level representations. This combination allows the network to attend to different aspects of the input, enhancing its ability to learn and represent diverse features.

The outputs of the Average Pooling and Max Pooling operations are then combined, often through element-wise addition or concatenation, to create the final output of the attention layer. This final output represents a fused representation that incorporates both average-level and maximum-level information, resulting in an attention effect that highlights relevant features in the data.

$$\begin{aligned} & \text{Attention}_{output} \\ & = \text{Concatenated}_{Features} \cdot \text{AveragePool}(\text{Concatenated}_{Features}) \\ & + \text{Concatenated}_{Features} \cdot \text{MaxPool}(\text{Concatenated}_{Features}) \end{aligned} \quad \dots(6)$$

where dot product represents element-wise multiplication.

$$\begin{aligned} & \text{ResidualBlock}_{output} \\ & = \text{ResidualPool}(\text{Attention}_{output}) \end{aligned} \quad \dots(7)$$

reducing dimensions to (3x3x4096).

$$\begin{aligned} & \text{Final}_{Features} \\ & = \text{Concatenate}(\text{ResidualBlock}_{output}, \text{Attention}_{output}) \end{aligned} \quad \dots(8)$$

combining the features from the Residual Pooling block and the Attention Layer.

$$\text{FinalBlock}_{output} = \text{ResidualBlock}(\text{Final}_{features}) \quad \dots(9)$$

further enhancing feature representation.

$$\text{GlobalPool}_{Output} = \text{GlobalAvgPool}(\text{FinalBlock}_{output}) \quad \dots(10)$$

resulting in a feature vector.

$$\text{Dense}_{output} = \text{ReLU}(\text{GlobalPool}_{output} \cdot \text{Weights} + \text{bias}) \quad \dots(11)$$

where Weights and bias are the learnable parameters of the Dense layer.

$$\text{Final}_{classification} = \text{Softmax}(\text{Dense}_{output}) \quad \dots(12)$$

assigning the features to one of the eight classes using the Softmax activation function.

IV. Results and Discussion

Analysis of proposed model is performed for 4 way classification of maize leaf disease detection. Table and Figure show that proposed model shows better results over ResNet101 and Xception Net with addition of attention layer. The performance parameters used for evaluation of the model are shown in table 6.

Table 6: Performance Parameters

Accuracy	TP+TN / (TP+TN+FP+FN)
Specificity	TN/(TN+FP)
Sensitivity	TP/(TP+FN)

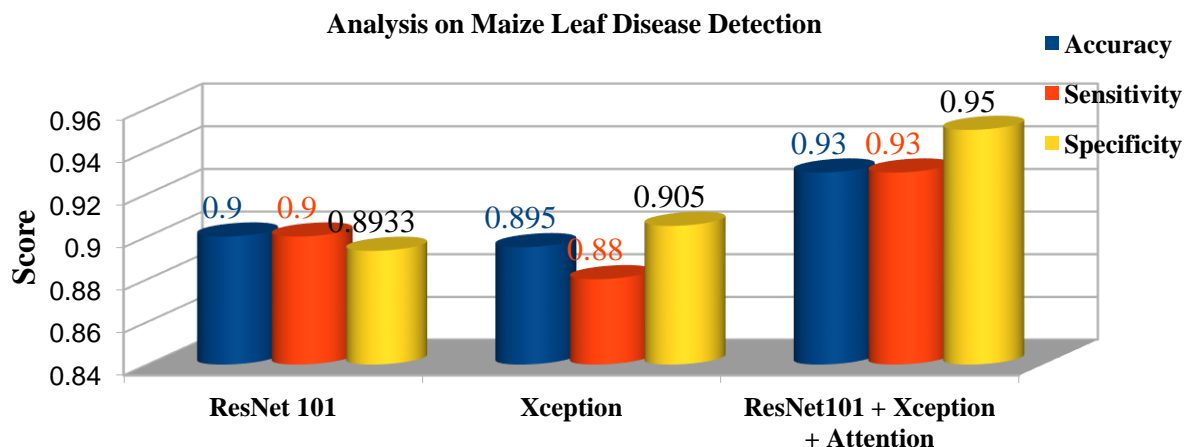


Figure 4: Analysis of proposed Model on Maize Leaf Disease Detection

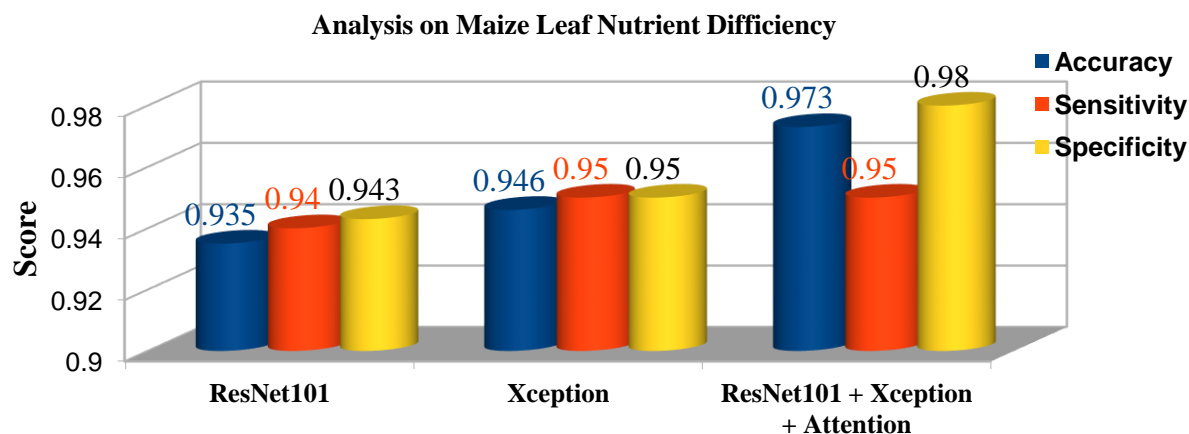


Figure 5: Analysis for Maize Leaf Nutrient Deficiency detection

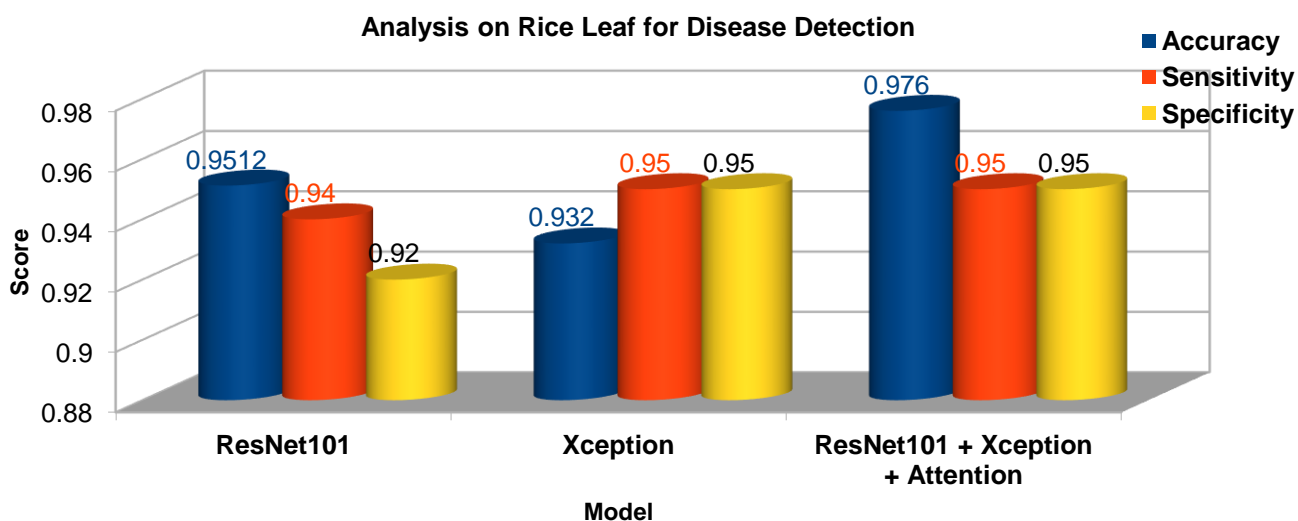


Figure 6: Analysis for Rice Leaf Disease detection

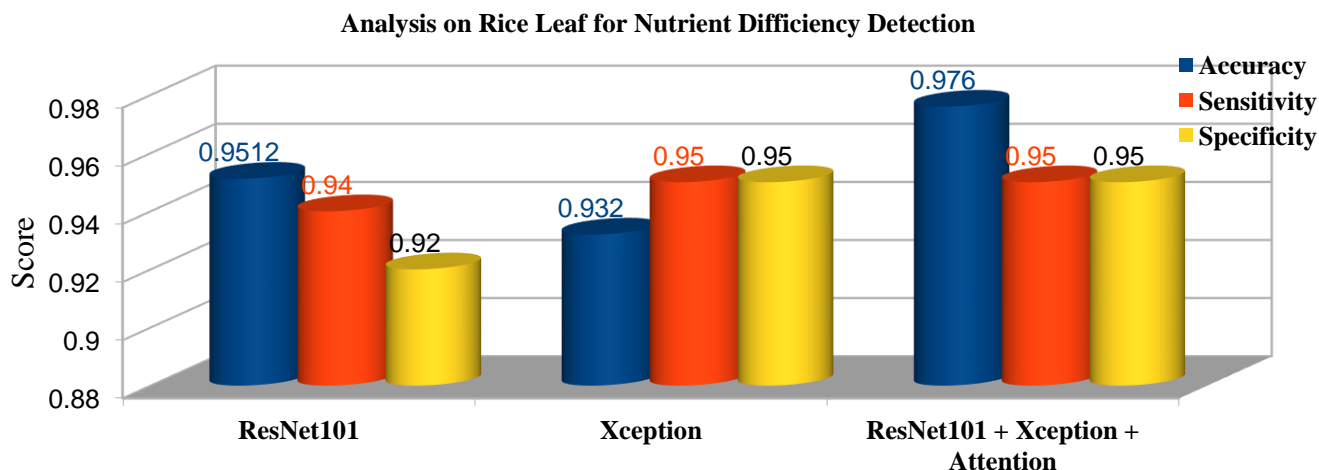


Figure 7: Analysis for Rice leaf nutrient deficiency detection

Comparative analysis:

In the existing methods, many researchers have considered maize and rice crops, one at a time. The disease detection work have included nutrient deficiency detection work. The comparative analysis of existing other methods with common crop factors are considered as shown Table 7.

Table 7: Comparative Analysis with other existing methods.

Method	Diseased Crop	Nutrient Deficiency Crop	Accuracy
Transfer Learning (VGG16,19, ResNet 18, GoogleNet) [31]	Maize	--	98%
Faster RCNN, YOLO-V2, RetinaNet [32]	Maize	--	95%
Convolutional Neural Network and OpenMP implementation [33]	Maize	--	93%
Custom CNN and Pretrained AlexNet [34]	Maize	--	97%
ResNet50 [35]	Rice	--	99%
GoogleNet, MobileNet-V3, AlexNet+SVM [36]	Rice	--	99%
Proposed	Maize	--	95%
Proposed	--	Maize	97.3%
Proposed	Rice	--	97.6%
Proposed	--	Rice	97.6%

Discussion:

The comparison of various methods for disease detection and nutrient deficiency identification in maize and rice crops is presented in Table 7. It is noteworthy that in the previous research, the focus was often directed towards analyzing either maize or rice crops individually, rather than considering both together. Moreover, the disease detection techniques were extended to include the identification of nutrient deficiencies in plants as well. This comprehensive evaluation sheds light on the performance of different methods in tackling these agricultural challenges. The first set of methods employs Transfer Learning techniques, utilizing architectures like VGG16, VGG19, ResNet 18, and GoogleNet. These methods were applied to detect diseases in maize crops, achieving an impressive accuracy of 98%. Following this, Faster RCNN, YOLO-V2, and RetinaNet were employed, again on maize crops, resulting in a slightly lower accuracy of 95%. Utilizing Convolutional Neural Networks (CNN) and OpenMP implementation led to a disease detection accuracy of 93% in maize. Custom CNN in combination with a pre-trained AlexNet yielded promising results with a disease detection accuracy of 97% in maize crops. Shifting the focus to rice crops, ResNet50 displayed remarkable performance with an accuracy of 99% in disease detection. GoogleNet, MobileNet-V3, and AlexNet with SVM achieved a similar accuracy of 99% in detecting diseases in rice crops. The proposed methods are introduced to enhance disease detection and nutrient deficiency identification. For maize crops, the proposed approach achieved an accuracy of 95%, while in the case of rice, it attained an accuracy of 97.6%. Interestingly, the proposed method targeting maize crops attained an accuracy of 97.3% even without specifying the nutrient deficiency factor. Similarly, the proposed method for rice crops, without considering nutrient deficiencies, achieved an accuracy of

97.6%. The fusion of Xception and ResNet101, along with the addition of an attention mechanism, can result in a highly complex model architecture. This complexity may require significant computational resources, including memory and processing power, which could limit its practicality for deployment on resource-constrained devices or in regions with limited access to advanced hardware.

V. Conclusion

In conclusion, the landscape of disease detection and nutrient deficiency identification in crops has seen significant advancements through various methodologies, as highlighted in the comparative analysis presented in Table 7. While individual methods have demonstrated commendable accuracy, the proposed model that integrates Xception and ResNet101, augmented by an attention mechanism, emerges as a potential solution to these challenges. However, this study also underscores several considerations and limitations that warrant attention. The proposed model's amalgamation of state-of-the-art architectures showcases its potential for enhanced performance in detecting diseases and nutrient deficiencies. Its utilization of both Xception and ResNet101, along with an attention mechanism, reflects a sophisticated approach to feature extraction and focus on crucial regions within images. This bodes well for accurate identification and diagnosis, essential for optimizing crop health and yield. Nonetheless, the model's complexity demands thorough evaluation. Balancing computational demands with accessibility to resources remains a pivotal concern, ensuring the model's applicability across diverse agricultural settings. Moreover, the potential lack of interpretability in deep models could hinder their adoption by end-users seeking insights into decision-making processes.

Furthermore, the model's effectiveness hinges upon robust training data and generalizability. Addressing biases and limitations in the dataset is imperative to avoid suboptimal performance and false predictions. The reliance on specific crops (maize and rice) necessitates caution when extrapolating results to other crops with distinct disease patterns. Despite these limitations, the proposed model presents a stepping stone towards holistic crop management. As agricultural systems grow more complex, incorporating diverse factors like pest infestations, soil health, and environmental conditions into future iterations of the model could yield more comprehensive insights. Furthermore, devising user-friendly interfaces and ensuring seamless integration into real-world farming practices will enhance the model's practical utility.

In essence, the proposed model's strengths in disease detection and nutrient deficiency identification underscore its potential to revolutionize agricultural practices. While challenges persist, an adaptive and iterative approach to model development and

deployment can pave the way for sustainable crop production, benefiting farmers and food security worldwide.

References:

- [1] L. Alzubaidi et al., "Review of deep learning: concepts, CNN architectures, challenges, applications, future directions," *J. Big Data* 2021 81, vol. 8, no. 1, pp. 1–74, Mar. 2021, doi: 10.1186/S40537-021-00444-8.
- [2] I. Buja et al., "Advances in Plant Disease Detection and Monitoring: From Traditional Assays to In-Field Diagnostics," *Sensors (Basel)*, vol. 21, no. 6, pp. 1–22, Mar. 2021, doi: 10.3390/S21062129.
- [3] M. M. Taye, "Theoretical Understanding of Convolutional Neural Network: Concepts, Architectures, Applications, Future Directions," *Comput.* 2023, Vol. 11, Page 52, vol. 11, no. 3, p. 52, Mar. 2023, doi: 10.3390/COMPUTATION11030052.
- [4] H. D. Mafukidze, G. Owomugisha, D. Otim, A. Nechibvute, C. Nyamhere, and F. Mazunga, "Adaptive Thresholding of CNN Features for Maize Leaf Disease Classification and Severity Estimation," *Appl. Sci.* 2022, Vol. 12, Page 8412, vol. 12, no. 17, p. 8412, Aug. 2022, doi: 10.3390/AP12178412.
- [5] N. Ahmad, H. M. S. Asif, G. Saleem, M. U. Younus, S. Anwar, and M. R. Anjum, "Leaf Image-based Plant Disease Identification using Color and Texture Features," *Wirel. Pers. Commun.*, vol. 121, no. 2, pp. 1139–1168, Feb. 2021, doi: 10.1007/s11277-021-09054-2.
- [6] A. Antunes, B. Ferreira, N. Marques, and N. Carriço, "Hyperparameter Optimization of a Convolutional Neural Network Model for Pipe Burst Location in Water Distribution Networks," *J. Imaging* 2023, Vol. 9, Page 68, vol. 9, no. 3, p. 68, Mar. 2023, doi: 10.3390/JIMAGING9030068.
- [7] K. He, X. Zhang, S. Ren, and J. Sun, "Deep Residual Learning for Image Recognition," *Proc. IEEE Comput. Soc. Conf. Comput. Vis. Pattern Recognit.*, vol. 2016-December, pp. 770–778, Dec. 2015, doi: 10.1109/CVPR.2016.90.
- [8] F. Chollet, "Xception: Deep Learning with Depthwise Separable Convolutions," *Proc. - 30th IEEE Conf. Comput. Vis. Pattern Recognition, CVPR 2017*, vol. 2017-January, pp. 1800–1807, Oct. 2016, doi: 10.1109/CVPR.2017.195.
- [9] A. Abade, P. A. Ferreira, and F. de Barros Vidal, "Plant Diseases recognition on images using Convolutional Neural Networks: A Systematic Review," *Comput. Electron. Agric.*, vol. 185, Sep. 2020, doi: 10.1016/j.compag.2021.106125.
- [10] V. S. Dhaka et al., "A Survey of Deep Convolutional Neural Networks Applied for Prediction of Plant Leaf Diseases," *Sensors* 2021, Vol. 21, Page 4749, vol. 21, no. 14, p. 4749, Jul. 2021, doi: 10.3390/S21144749.
- [11] M. Nagaraju and P. Chawla, "Systematic review of deep learning techniques in plant disease detection," *Int. J. Syst. Assur. Eng. Manag.*, vol. 11, no. 3, pp. 547–560, Jun. 2020, doi: 10.1007/S13198-020-00972-1/FIGURES/9.
- [12] A. Kamilaris and F. X. Prenafeta-Boldú, "Deep learning in agriculture: A survey," *Comput. Electron. Agric.*, vol. 147, pp. 70–90, Apr. 2018, doi:

- 10.1016/J.COMPAG.2018.02.016.
- [13] C. Fernández-Quintanilla, J. M. Peña, D. Andújar, J. Dorado, A. Ribeiro, and F. López-Granados, "Is the current state of the art of weed monitoring suitable for site-specific weed management in arable crops?," *Weed Res.*, vol. 58, no. 4, pp. 259–272, Aug. 2018, doi: 10.1111/WRE.12307.
- [14] W. Setiawan, E. M. S. Rochman, B. D. Satoto, and A. Rachmad, "Machine Learning and Deep Learning for Maize Leaf Disease Classification: A Review," *J. Phys. Conf. Ser.*, vol. 2406, no. 1, p. 012019, Dec. 2022, doi: 10.1088/1742-6596/2406/1/012019.
- [15] J. Lu, L. Tan, and H. Jiang, "Review on Convolutional Neural Network (CNN) Applied to Plant Leaf Disease Classification," *Agric.* 2021, Vol. 11, Page 707, vol. 11, no. 8, p. 707, Jul. 2021, doi: 10.3390/AGRICULTURE11080707.
- [16] K. Golhani, S. K. Balasundram, G. Vadamalai, and B. Pradhan, "A review of neural networks in plant disease detection using hyperspectral data," *Inf. Process. Agric.*, vol. 5, no. 3, pp. 354–371, Sep. 2018, doi: 10.1016/J.INPA.2018.05.002.
- [17] M. H. Al-Adhaileh, A. Verma, T. H. H. Aldhyani, and D. Koundal, "Potato Blight Detection Using Fine-Tuned CNN Architecture," *Math.* 2023, Vol. 11, Page 1516, vol. 11, no. 6, p. 1516, Mar. 2023, doi: 10.3390/MATH11061516.
- [18] G. Huang, Z. Liu, L. Van Der Maaten, and K. Q. Weinberger, "Densely Connected Convolutional Networks," *Proc. - 30th IEEE Conf. Comput. Vis. Pattern Recognition, CVPR 2017*, vol. 2017-Janua, pp. 2261–2269, Aug. 2016, doi: 10.1109/CVPR.2017.243.
- [19] W. Li et al., "Fraxin ameliorates lipopolysaccharide-induced acute lung injury in mice by inhibiting the NF- κ B and NLRP3 signalling pathways," *Int. Immunopharmacol.*, vol. 67, pp. 1–12, Feb. 2019, doi: 10.1016/J.INTIMP.2018.12.003.
- [20] S. H. Lee, H. Gočau, P. Bonnet, and A. Joly, "New perspectives on plant disease characterization based on deep learning," *Comput. Electron. Agric.*, vol. 170, p. 105220, Mar. 2020, doi: 10.1016/J.COMPAG.2020.105220.
- [21] Y. Mao, Z. He, Z. Ma, X. Tang, and Z. Wang, "Efficient Convolution Neural Networks for Object Tracking Using Separable Convolution and Filter Pruning," *IEEE Access*, vol. 7, pp. 106466–106474, 2019, doi: 10.1109/ACCESS.2019.2932733.
- [22] P. Singh, V. K. Verma, P. Rai, and V. P. Nambodiri, "Acceleration of Deep Convolutional Neural Networks Using Adaptive Filter Pruning," *IEEE J. Sel. Top. Signal Process.*, vol. 14, no. 4, pp. 838–847, May 2020, doi: 10.1109/JSTSP.2020.2992390.
- [23] G. Li, J. Wang, H. W. Shen, K. Chen, G. Shan, and Z. Lu, "CNNPruner: Pruning Convolutional Neural Networks with Visual Analytics," *IEEE Trans. Vis. Comput. Graph.*, vol. 27, no. 2, pp. 1364–1373, Sep. 2020, doi: 10.1109/TVCG.2020.3030461.
- [24] Z. Ma et al., "Maize leaf disease identification using deep transfer convolutional neural networks," *Int. J. Agric. Biol. Eng.*, vol. 15, no. 5, pp. 187–195, Nov. 2022, doi: 10.25165/IJABE.V15I5.6658.
- [25] P. Rawat, A. Pandey, and A. Panaiyappan.k, "Rice Leaf Diseases Classification Using Deep Learning Techniques," *Proc. 1st IEEE Int. Conf. Netw. Commun. 2023, ICNWC 2023*, 2023, doi: 10.1109/ICNWC57852.2023.10127315.
- [26] L. Yang et al., "Stacking-based and improved convolutional neural network: a new approach in rice leaf disease identification," *Front. Plant Sci.*, vol. 14, p. 1165940, Jun. 2023, doi: 10.3389/FPLS.2023.1165940/BIBTEX.
- [27] H. Yang et al., "Automatic Recognition of Rice Leaf Diseases Using Transfer Learning," *Agron.* 2023, Vol. 13, Page 961, vol. 13, no. 4, p. 961, Mar. 2023, doi: 10.3390/AGRONOMY13040961.
- [28] S. M. Javidan, A. Banakar, K. A. Vakilian, and Y. Ampatzidis, "Tomato leaf diseases classification using image processing and weighted ensemble learning," *Agron. J.*, 2023, doi: 10.1002/AGJ2.21293.
- [29] A. P. J and G. GOPAL, "Data for: Identification of Plant Leaf Diseases Using a 9-layer Deep Convolutional Neural Network," vol. 1, 2019, doi: 10.17632/TYWBTJRJV.1.
- [30] H. B. Prajapati, J. P. Shah, and V. K. Dabhi, "Detection and classification of rice plant diseases," *Intell. Decis. Technol.*, vol. 11, no. 3, pp. 357–373, Jan. 2017, doi: 10.3233/IDT-170301.
- [31] R. Hu, S. Zhang, P. Wang, G. Xu, D. Wang, and Y. Qian, "The identification of corn leaf diseases based on transfer learning and data augmentation," *ACM Int. Conf. Proceeding Ser.*, pp. 58–65, May 2020, doi: 10.1145/3403746.3403905.
- [32] A. Hamidisepehr, S. V. Mirmezami, and J. K. Ward, "COMPARISON OF OBJECT DETECTION METHODS FOR CORN DAMAGE ASSESSMENT USING DEEP LEARNING."
- [33] D. A. Padilla, R. A. I. Pajes, and J. T. De Guzman, "Detection of Corn Leaf Diseases Using Convolutional Neural Network with OpenMP Implementation," *2020 IEEE 12th Int. Conf. Humanoid, Nanotechnology, Inf. Technol. Commun. Control. Environ. Manag. HNICEM 2020*, Dec. 2020, doi: 10.1109/HNICEM51456.2020.9400004.
- [34] P. K. Singamsetty, G. V. N. D. S. Prasad, N. V. S. Naidu, and R. S. Kumar, "Maize Leaf Disease Detection and Classification Using Deep Learning," *Artif. Intell. Mech. Ind. Eng.*, pp. 87–102, Jun. 2021, doi: 10.1201/9781003011248-6.
- [35] M. S. A. M. Al-Gaashani, N. A. Samee, R. Alnashwan, M. Khayyat, and M. S. A. Muthanna, "Using a Resnet50 with a Kernel Attention Mechanism for Rice Disease Diagnosis," *Life*, vol. 13, no. 6, p. 1277, May 2023, doi: 10.3390/LIFE13061277.
- [36] M. H. Saleem, J. Potgieter, and K. M. Arif, "Plant Disease Detection and Classification by Deep Learning," *Plants* 2019, Vol. 8, Page 468, vol. 8, no. 11, p. 468, Oct. 2019, doi: 10.3390/PLANTS8110468.